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Molecular Pathogenesis of Polymerase Gamma–Related Neurodegeneration

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Objective: Polymerase gamma (POLG) mutations are a common cause of mitochondrial disease and have also been linked to neurodegeneration and aging. We studied the molecular mechanisms underlying POLG-related neurodegeneration using postmortem tissue from a large number of patients.

Methods: Clinical information was available from all subjects. Formalin-fixed and frozen brain tissue from 15 patients and 23 controls was studied employing a combination of histopathology, immunohistochemistry, and molecular studies of microdissected neurons.

Results: The primary consequence of POLG mutation in neurons is mitochondrial DNA depletion. This was already present in infants with little evidence of neuronal loss or mitochondrial dysfunction. With longer disease duration, we found an additional, progressive accumulation of mitochondrial DNA deletions and point mutations accompanied by increasing numbers of complex I–deficient neurons. Progressive neurodegeneration primarily affected the cerebellar systems and dopaminergic cells of the substantia nigra. Superimposed on this chronic process were acute, focal cortical lesions that correlated with epileptogenic foci and that showed massive neuronal loss.

Interpretation: POLG mutations appear to compromise neuronal respiration via a combination of early and stable depletion and a progressive somatic mutagenesis of the mitochondrial genome. This leads to 2 distinct but overlapping biological processes: a chronic neurodegeneration reflected clinically by progressive ataxia and cognitive impairment, and an acute focal neuronal necrosis that appears to be related to the presence of epileptic seizures. Our findings offer an explanation of the acute-on-chronic clinical course of this common mitochondrial encephalopathy.
accurately reflect human POLG encephalopathy syndromes. It is known that POLG mutations affect the enzyme’s ability to efficiently replicate and repair mtDNA,10,11 and patients with POLG disease do harbor mtDNA damage and signs of respiratory chain dysfunction in affected tissues.12–15 The nature of the mtDNA defect appears to differ depending on which tissue is studied,1,12,16 with the central nervous system (CNS) being least well understood due to the difficulties in obtaining human tissues to study.

The few human studies of neuronal mtDNA in POLG disease have shown inconsistent results. Multiple mtDNA deletions affecting between ~20 and 50% of mtDNA molecules have been identified,12–15 but whether this level of mtDNA deletion alone is sufficient to cause respiratory dysfunction and neuronal death remains unclear.17 Both normal and mildly decreased (70–75% of control) mtDNA copy number have been found in brain homogenate,12,13,16,18 and 1 study in neurons from the dorsal root ganglia of a single patient15 has reported clear mtDNA depletion (~50% of controls). Accumulation of mtDNA point mutations has been shown in skeletal muscle of patients with POLG disease using an ultradepth resequencing-by-synthesis (UDS) assay,19,20 whereas no increase in mtDNA point mutations was detected in another study in brain.12 This study used a strategy of polymerase chain reaction (PCR) and cloning that has limited sensitivity, however, and cannot exclude mutations present at low heteroplasmy levels.

POLG disease affecting the brain often manifests as an acute-on-chronic process.8,9 To understand this at the level of the neuron, we investigated the changes in respiratory chain proteins and mtDNA in extensive material comprising tissue from all major areas of the CNS from 15 patients aged 0.6 to years 53 years with encephalopathy syndromes caused by the most common POLG mutations (c.1399G>A, p.A467T or c.2243G>C, p.W748S). Our findings show that it is possible to construct a sequence of events linking early quantitative loss of mtDNA and subsequent accumulation of mtDNA damage to respiratory chain deficiency and neuronal injury.

Subjects and Methods

Patients and Controls
Postmortem CNS tissue from 15 patients with POLG encephalopathy and 23 controls was studied. Demographic data and the tissues and studies that were performed are summarized in the Supplementary Table.

The patients all had a progressive MSCAE with focal and secondary generalized seizures and suffered at least 1 stroke-like episode with cortical stroke-like lesions on magnetic resonance imaging (MRI). In 12 patients for whom electroencephalographic (EEG) data were available, the localization of the cortical lesions correlated with epileptiform activity. Three patients had infantile onset encephalopathy with severe epilepsy and stroke-like episodes. The clinical and genetic features of all patients are summarized in Table 1 and have been described elsewhere in detail.8,9

Controls had no known neurological disease or injury, did not use antiviral medication, and had normal routine histology in all brain samples collected. Fresh frozen tissue was collected prospectively from 17 controls, comprising 12 adults and children with a mean age of 42.3 ± 22.2 years and 5 infants (<1 year old) with a mean age of 0.11 ± 0.16 years; formalin-fixed paraffin-embedded (FFPE) tissue was available from a further 6 subjects with an average age of 42 ± 9.7 years and an 85-year-old patient with advanced, histologically confirmed Alzheimer disease.

Tissue Material
Samples were collected from the frontal, temporal, and occipital cortex, hippocampus, thalamus, striatum (putamen), cerebellum, dentate nucleus, mesencephalon at the level of the substantia nigra, pons, medulla oblongata at the level of the inferior olivary nucleus, and cervical spinal cord. Samples were dissected at autopsy and either snap-frozen in isopentane cooled in liquid nitrogen and stored at −80°C or fixed in formaldehyde and embedded in paraffin blocks according to standard procedures. There were no significant differences in postmortem interval or length of fixation between patient and control tissue.

Histology and Morphometric Analysis
Routine investigation of FFPE sections included hematoxylin and eosin (H&E), cresyl violet, Luxol myelin stain, and glial fibrillar acidic protein immunohistochemistry. Routine morphological examination with H&E was performed on all samples from all patients and controls. Morphometric analysis and cell counting were performed using a light microscope (Leica, Wetzlar, Germany) equipped with a Zeiss AxioCam MRc5 camera and Zen 2011 software (Carl Zeiss MicroImaging, Jena, Germany). Cell counting was performed on 4-μm-thick sections immunohistochemically stained for porin (which stains all neurons irrespective of respiratory chain status) in the cerebellar cortex, dentate nucleus, and inferior olivary nucleus. Only neurons with a clearly visible nuclear morphology were counted. In the cerebellum, the length of the Purkinje cell layer was measured at ×25 magnification, all Purkinje cells fulfilling the above criteria were counted, and cell density was calculated as Purkinje cells/mm. The total area of the dentate and inferior olivary nuclei available on the section was measured, neurons were counted as above, and neuronal density was calculated as neurons/mm². Eosinophilic neuronal necrosis was defined as neurons with intensely eosinophilic cytoplasm and pyknotic nucleus with no discernible nucleolus.

Histochemistry and Immunohistochemistry
Immunohistochemistry for respiratory complexes I, II, III, and IV and the mitochondrial membrane protein porin was performed on 15 patients and histochemistry for cytochrome oxidase (COX) and succinate dehydrogenase (SDH) on 8 as previously described.6
<table>
<thead>
<tr>
<th>Patient</th>
<th>Mutation</th>
<th>Age at Onset, yr</th>
<th>Age at Death, yr</th>
<th>Epilepsy</th>
<th>SLE</th>
<th>Ataxia</th>
<th>Neuropathy</th>
<th>PEO</th>
<th>FFPE Tissue</th>
<th>Frozen Tissue</th>
</tr>
</thead>
<tbody>
<tr>
<td>AT-1A</td>
<td>A467T/A467T</td>
<td>15</td>
<td>44</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>AT-1B</td>
<td>A467T/A467T</td>
<td>8</td>
<td>47</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>-</td>
</tr>
<tr>
<td>AT-2A</td>
<td>A467T/A467T</td>
<td>16</td>
<td>53</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
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<td>+</td>
</tr>
<tr>
<td>WS-1A</td>
<td>W748S/W748S</td>
<td>6</td>
<td>41</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>WS-2A</td>
<td>W748S/W748S</td>
<td>15</td>
<td>24</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>WS-3A</td>
<td>W748S/W748S</td>
<td>17</td>
<td>43</td>
<td>+</td>
<td>+</td>
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</tr>
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<td>WS-8A</td>
<td>W748S/W748S</td>
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<td>28</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
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<tr>
<td>WS-9A</td>
<td>W748S/W748S</td>
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<td>13</td>
<td>+</td>
<td>+</td>
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<td>+</td>
<td>+</td>
<td>-</td>
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</tr>
<tr>
<td>WS-10A</td>
<td>W748S/W748S</td>
<td>16</td>
<td>24</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>WS-12A</td>
<td>W748S/W748S</td>
<td>15</td>
<td>57</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>+</td>
</tr>
<tr>
<td>CP-4A</td>
<td>A467T/W748S</td>
<td>14</td>
<td>23</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>CP-4B</td>
<td>A467T/W748S</td>
<td>13</td>
<td>21</td>
<td>+</td>
<td>+</td>
<td>+</td>
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<td>-</td>
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</tr>
<tr>
<td>Mean/total</td>
<td></td>
<td>12.2 ± 4.7</td>
<td>34.8 ± 14.3</td>
<td>12/12</td>
<td>12/12</td>
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<td>12/12</td>
<td>7/12</td>
<td>10/12</td>
</tr>
<tr>
<td>AL-1A</td>
<td>A467T/G303R</td>
<td>0.9</td>
<td>1.1</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>AL-1B</td>
<td>A467T/G303R</td>
<td>2.0</td>
<td>8.0</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>AL-2A</td>
<td>A467T/G848S</td>
<td>0.6</td>
<td>0.6</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>-</td>
</tr>
<tr>
<td>Mean/total</td>
<td></td>
<td>1.2 ± 0.7</td>
<td>3.2 ± 4.1</td>
<td>3/3</td>
<td>3/3</td>
<td>1/3</td>
<td>0/3</td>
<td>0/3</td>
<td>3/3</td>
<td>2/3</td>
</tr>
</tbody>
</table>

+ = feature present or tissue available; − = feature not present or tissue not available; FFPE = formalin-fixed, paraffin-embedded; PEO = progressive external ophthalmoplegia; SLE = stroke-like episode.
**Cell Death Studies**

Ten patients and 7 controls were studied (see Supplementary Table) by caspase-3 immunohistochemistry and terminal deoxynucleotidyl transferase–mediated deoxyuridine triphosphate nick-end labeling (TUNEL) assay. Caspase-3 immunohistochemistry was performed as previously described. TUNEL was performed on 8µm-thick frozen sections using a commercial kit (In Situ Cell Death Detection Kit, Fluorescein, v16.0; Roche, Basel, Switzerland) according to the manufacturer’s protocol.

**Neuron Microdissection**

Neurons for mtDNA analysis were microdissected from different CNS areas of 8 patients and 17 age-matched controls (see Supplementary Table). Microdissection and cell lysis were carried out as previously described. Only cells that could be positively identified as neurons, with a visible nucleus and normal morphological characteristics, were used. For each area, there was no significant size difference between neurons of patients and corresponding controls. A total of 4,012 neurons were picked from 8 patients (n = 1,896) and 17 age-matched controls (n = 2,116). Neurons were microdissected, avoiding carryover of glia or other cells, and pooled in groups of 10 in the cortex (n = 2,456 pyramidal neurons in total) and hippocampal CA formation (n = 250); 15 in the substantia nigra (n = 360); 5 in the cerebellum (n = 490 Purkinje cells), dentate (n = 165 principal dentate neurons), and olivary (n = 175 principal olivary neurons) nuclei; and 4 in the anterior horn of the spinal cord (n = 116 alpha motor neurons).

**mtDNA Studies**

mtDNA quantification and deletion assessment was performed in gray matter homogenate and in microdissected neurons of 8 patients and 17 age-matched controls (see Supplementary Table) using quantitative PCR (qPCR) and long-range PCR (LPCR) as previously described. Assessment of mtDNA point sequence variants was performed in genomic DNA extracted from frontal cortical homogenate of 6 patients (2 A467T homozygous and 4 W748S homozygous) and 5 age-matched controls (see Supplementary Table). Point mutations were detected with an UDS assay using FLX GS technology (Roche 454 Life Sciences, Branford, CT) as previously described. Two regions of mtDNA were amplified and subsequently sequenced: 1 in the noncoding hypervariable region MT-HV2, known to have a high mutational rate, and 1 in the highly conserved, peptide-coding MT-COβ gene. PCR products were purified using Agencourt Ampure XP beads (Beckman Coulter, Fullerton, CA) and quantified using a Qubit 2.0 Fluorometer (Life Technologies, Grand Island, NY). Samples were pooled into equimolar libraries and processed for bidirectional amplification sequencing using Titanium (XLR70) chemistry (Roche) and a GS-FLX (v2.8 sequencing software) according to the manufacturer’s instructions (Roche).

**Bioinformatic Analyses**

PyroBayes (v1.1) and Mosaik (v0.9.0891) algorithms were used in sequence base calling and alignment. Subsequent analysis of variants was done within R using a custom-made R library flowgram. Homopolymer tracts (n ≥ 4) within the amplicons were excluded from analysis, to minimize pyrosequencing derived base-calling error. Variants were recorded if present in both directions with ≥0.2% heteroplasmy as previously described.

**Statistical Analyses**

Statistical analyses were performed in SPSS (v.20.0.0.1; IBM, Armonk, NY) and Prism (v6; GraphPad, San Diego, CA). Comparison of proportions of point mutations between groups was done by chi-square test. Comparison of mtDNA quantity between groups was performed by Mann–Whitney U test. Cell densities were compared by 2-tailed t test, and correlation analysis was done using Pearson coefficient.

**Results**

**POLG Mutations Cause Acute Focal Neuronal Necrosis Superimposed on a Background of Chronic Neurodegeneration**

Histopathological findings fell into 3 categories: acute focal lesions, diffuse progressive changes with uniform intraregional distribution, and mildly or unaffected areas. These findings are detailed in Table 2.

Focal, sharply demarcated lesions were seen in the neocortex (14 of 14 patients), hippocampus CA region (4 of 14), thalamus (2 of 9), and cerebellar cortex (9 of 14). These were characterized by selective neuronal loss, eosinophilic neuronal necrosis, vacuolation of the neuropil, astrocytosis, and diffuse microglial activation (Fig 1, Table 2). Neuronal loss in the cerebral cortex had a laminar pattern, being most pronounced in the superficial (II–III) and deep (V–VI) layers. Surviving neurons with normal cytoplasmic and nuclear morphology were scattered throughout the lesion (see Fig 1E, F), and there was no infiltration by peripheral inflammatory cells. The cortical microvasculature in and around the lesions was patent, and in 4 cases increased vascularity was found within the lesions.

Cortical acute lesions correlated with the localization of acute strokelike lesions on MRI and epileptiform activity on EEG, and there was a clear predilection for the occipital and frontal lobes, whereas the temporal neocortex was universally spared (histologically and on MRI). All 4 patients with the earliest disease onset and age of death had focal lesions confined to the CA1 area of the hippocampus, whereas the hippocampal formation showed normal morphology in the remaining patients. Focal lesions were seen in the cerebellar cortex of 8 patients and were characterized by severe Purkinje and granule cell loss and Bergmann gliosis. These lesions could be multiple, but were often microscopic (Fig 2E, F) and not always detected by MRI.
TABLE 2. Pathology Findings in Polymerase Gamma Encephalopathy

<table>
<thead>
<tr>
<th>Patient</th>
<th>AAD</th>
<th>FENN Lesion</th>
<th>Neuronal Loss and Gliosis</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Cerebral Cortex</td>
<td>Cerebellar Cortex</td>
</tr>
<tr>
<td>AT-1A</td>
<td>44</td>
<td>Y</td>
<td>N</td>
</tr>
<tr>
<td>AT-1B</td>
<td>47</td>
<td>Y</td>
<td>N</td>
</tr>
<tr>
<td>AT-2A</td>
<td>53</td>
<td>Y</td>
<td>Y</td>
</tr>
<tr>
<td>WS-1A</td>
<td>41</td>
<td>Y</td>
<td>N</td>
</tr>
<tr>
<td>WS-3A</td>
<td>43</td>
<td>Y</td>
<td>N</td>
</tr>
<tr>
<td>WS-8A</td>
<td>28</td>
<td>Y</td>
<td>N</td>
</tr>
<tr>
<td>WS-9A</td>
<td>13</td>
<td>Y</td>
<td>Y</td>
</tr>
<tr>
<td>WS-10A</td>
<td>24</td>
<td>Y</td>
<td>N</td>
</tr>
<tr>
<td>WS-12A</td>
<td>57</td>
<td>Y</td>
<td>Y</td>
</tr>
<tr>
<td>CP-4A</td>
<td>23</td>
<td>Y</td>
<td>N</td>
</tr>
<tr>
<td>CP-4B</td>
<td>21</td>
<td>Y</td>
<td>Y</td>
</tr>
<tr>
<td>AL-1A</td>
<td>1.1</td>
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<td>N</td>
</tr>
<tr>
<td>AL-1B</td>
<td>8.0</td>
<td>Y</td>
<td>Y</td>
</tr>
<tr>
<td>AL-2A</td>
<td>0.6</td>
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<td>N</td>
</tr>
<tr>
<td>Total</td>
<td>14/14</td>
<td>9/14</td>
<td>9/9</td>
</tr>
</tbody>
</table>

Pathology is graded according to severity as normal (–), mild (+), moderate (++), and severe (+++). AAD = age at death in years; DCD = dorsal column degeneration in the spinal cord; FENN = focal energy-dependent neuronal necrosis; N = absent; NA = not available; Y = present.
FIGURE 1: Histopathological and immunohistochemical analysis of acute lesions in polymerase gamma encephalopathy. Brain histology from a representative patient (WS-1A) compares unaffected cortex (left side, A–D) and an acute cortical lesion (right side, E–I). Sections are either stained with hematoxylin and eosin (A, E, F), or react immunohistochemically to glial fibrillary acidic protein (B, G), the microglial marker HLA-DR (C, H), or complex I subunit NDUFB8 (D, I). Original magnification = ×100. (F) Magnified (original magnification = ×400) detail from E. The acute lesion is characterized by severe but incomplete neuronal loss (E and F) and vacuolation of the neuropil. Surviving neurons with normal morphological characteristics (arrows in F) are scattered throughout the acute lesion. There is pronounced astrocytosis (G) and diffuse microglial activation (H). In the neighboring, morphologically preserved cortex (D, arrows), there is a high proportion of complex I-negative neurons, but only complex I-positive cells survive within the acute lesion (I). Scale bars = 100μm (A–C, E, G, H), 50μm (D, F, I).
Diffuse, uniformly distributed changes consisted of neuronal loss and gliosis affecting mainly the cerebellar dentato-olivary and dentatorubral systems, thalamus, and substantia nigra. In the cerebellar cortex, there was diffuse, uniform Purkinje cell loss in addition to the focal lesions (see Fig 2). The dentate nucleus showed severe neuronal loss in addition to pronounced cytoplasmic accumulation of lipofuscin pigment in remaining neurons. Purkinje cell loss was significantly ($p = 0.0006$) more severe in patients homozygous for W748S and compound heterozygous patients (Purkinje cell count = 29.2 ± 14.1% of controls) than in the A467T homozygous patients, who showed only mild reduction of Purkinje cell numbers (Purkinje cell count = 81.5 ± 13.4% of controls). Neuronal loss in the dentate also appeared to be more pronounced in the W748S and compound heterozygous patients (dentate neuronal count = 47.7 ± 28.5% of controls) than in the A467T homozygous patients (dentate neuronal count = 82.4 ± 22.6% of controls), but this difference was not statistically significant. Cerebellar neuronal loss correlated with MRI findings of more pronounced cerebellar and dentate atrophy in the W748S homozygous and compound heterozygous patients than in the A467T homozygous patients. Despite this, we detected no significant clinical difference in the severity of ataxia in the 2 groups.

**FIGURE 2:** Cerebellar pathology in polymerase gamma encephalopathy: comparison of patients with the W748S and/or A467T mutations. Cerebellar sections from patients stained with hematoxylin and eosin (A–D) or porin immunohistochemistry (E, F) are shown. Patients are homozygous for the A467T (A, B), homozygous for the W748S (C, D), or compound heterozygous in transit for the A467T and W748S mutations (E, F). The cerebellum shows a combination of 2 types of pathology: diffuse neurodegeneration with Purkinje cell loss, Bergmann gliosis, and thinning of the molecular layer (A–F) and microscopic focal, sharply demarcated lesions (E, F). Diffuse degenerative changes are significantly more severe in the W748S homozygous (C, D) and compound heterozygous (E, F) compared to A467T homozygous (A, B) patients.
The thalamus was abnormal in all patients studied (9 of 9), showing diffuse neuronal loss and gliosis in addition to circumscribed focal lesions in 2 patients. In the substantia nigra, there was severe loss of pigmented, dopaminergic neurons, which was most pronounced in the ventrolateral tier. A detailed description of the nigral pathology is described elsewhere. In the brainstem, there was pronounced neuronal loss and gliosis in the locus coeruleus and the inferior olivary nucleus. In the spinal cord, there was severe, selective degeneration of the dorsal funiculus, which was more pronounced in the fasciculus gracilis, as previously described.

Regions of the cerebral neocortex and hippocampus not affected by focal lesions and the striatum showed either normal morphology or only mild neuronal loss as evidenced by mildly increased numbers of astrocytes in the cortex and underlying white matter. The pons and medulla oblongata appeared structurally preserved, with the exception of the locus coeruleus and olivary nucleus.

Patient Neurons Show Selective and Progressively Increasing Deficiency of Respiratory Complex I

Immunohistochemistry revealed significant numbers of complex I–deficient neurons in POLG patients (Table 3, Fig 3), but no deficiency of complexes II and III or porin and only a small number of complex IV–deficient neurons identified either by COX/SDH histochemistry or immunohistochemistry. No respiratory chain–deficient neurons were found in the controls, with the exception of a few complex I–negative neurons in the substantia nigra (12.5%) and the CA area of the hippocampus (<1%) of the patient with Alzheimer disease.

The numbers of neurons negative for complex I and IV varied according to region and which type of pathological process affected the area under scrutiny. In focal, acute lesions, with few remaining neurons, most surviving neurons retained complex I staining (see Fig 1I). Thus, the proportion of complex I–negative neurons was significantly lower in these areas (p = 0.03) than in those affected by the chronic process. Acute lesions also contained no complex IV–negative cells. In regions with better-preserved neuronal populations, that is, chronically affected, there were abundant complex I–deficient neurons, and the relative proportion of deficient to nondeficient neurons was accordingly higher (see Table 3, Figs 1D and 3A). Moreover, in these regions with surviving neurons, the number of neurons deficient for complex I (and IV) increased significantly with patient age (see Fig 3J). Few complex I–negative neurons were found in the infant patients, with the exception of the substantia nigra, which showed ~60% complex I–negative neurons already at the age of 1 year (Patient AL-2A).

The most severe complex I (and IV) defects were found in the substantia nigra, red nucleus, olivary nucleus, and central basal pontine nuclei (peduncular and median), followed by the dentate nucleus, cerebellar Purkinje cells, hippocampal CA2 and CA4 areas, and cerebral neocortex (see Table 3). There was no significant difference in the numbers of neurons negative for complex I or IV between the frontal, temporal, and occipital cortices. The mean ratio of complex IV–negative to complex I–negative neurons showed regional variation being substantially higher in the cerebral cortex (0.35) and Purkinje cells (0.23) than in deeper nuclei, including the dentate (0.03), inferior olive (0.008), and substantia nigra (0.02).

Studies of Cell Death

Caspase-3 staining revealed no positive nuclei in any of the patients. TUNEL was negative in the adult patients, whereas several positive nuclei were seen in Patients AL-1A (age = ~1 year) and AL-1B (age = 8 years). The same areas were, however, negative for caspase-3.

POLG Mutations Cause Early Quantitative Depletion and Progressive Accumulation of Somatic Damage in the Mitochondrial Genome

Quantification of mtDNA in gray matter homogenate showed normal (n = 5) or moderately reduced (n = 2) mtDNA levels in adult patients and apparent depletion (~20–30% of age-matched controls) in the children (Patients AL-1A and AL-1B). Investigation of microdissected neurons, however, revealed a different picture. Patient neurons consistently showed a substantial mtDNA depletion compared with age-matched control microdissected neurons; patients had approximately 40 ± 11% (range = 17–56%) of control mtDNA. This difference was found throughout the CNS, with no regional differences, and was highly significant (Figs 4 and 5C). Infants (≤1 year old) had a significantly lower neuronal mtDNA copy number than older individuals, in both the patients and controls. Neuronal mtDNA levels of postinfant and adult patients did not exceed those of control infants (see Fig 4).

Deletions of mtDNA were detectable by LPCR in gray matter homogenate and single neurons of patients. Deletions appeared as smears on homogenate PCR, whereas single neurons contained ≥1 deleted species (see Fig 5). Relative quantification of deleted mtDNA by qPCR in microdissected neurons showed that deletion levels in the patients ranged from undetectable in most CNS areas of the infant (Patient AL-1A) and child...
<table>
<thead>
<tr>
<th>Patient</th>
<th>AAD</th>
<th>Frontal Cortex</th>
<th>Hippocampus</th>
<th>Subiculum</th>
<th>Purkinje Cells</th>
<th>Dentate Nucleus</th>
<th>Olivary Nucleus</th>
<th>Substantia Nigra</th>
<th>Red Nucleus</th>
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<tr>
<td>AT-1A</td>
<td>44</td>
<td>55.6</td>
<td>NA</td>
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<td>NA</td>
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<tr>
<td>AT-1B</td>
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<td>58</td>
<td>9.5</td>
<td>63.7</td>
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<td>9.9</td>
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<tr>
<td>AT-2A</td>
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<tr>
<td>WS-1A</td>
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<td>16</td>
<td>82.5</td>
<td>30.5</td>
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<td>34.2</td>
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<td>18.9</td>
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<tr>
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<tr>
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<td>NA</td>
<td>NA</td>
<td>0</td>
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<td>0</td>
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<tr>
<td>AL-2A</td>
<td>0.65</td>
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<td>0</td>
<td>58.3</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>4.3</td>
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<tr>
<td>Mean</td>
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<td>0</td>
<td>45.0</td>
<td>0.7</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
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<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

AAD = age at death in years; DG = dentate gyrus; NA = not available; STD = standard deviation.
FIGURE 3: Respiratory chain dysfunction in polymerase gamma (POLG) encephalopathy. Serial sections of the hippocampal CA2 region (A–F) and inferior olivary nucleus (G–I) from a representative patient (AT-1B) are shown. There is severe and selective complex I (anti-NDUFB8) deficiency (A, G), with only a few complex IV–negative neurons in the hippocampus (D) and none in the olive (H), and normal staining for complexes II (B), III (C), and porin (E, I). Complex I stained all neurons in control hippocampus (F). Neuronal complex I deficiency is progressive in POLG encephalopathy (J). The proportion of complex I–negative neurons increased with age in all central nervous system areas studied. The diagrams in J show the percentage of complex I–negative neurons (y-axis) as a function of individual patient age in years (x-axis) in the frontal cortex, hippocampal CA2 region, substantia nigra, and anterior spinal horn. The dopaminergic neurons of the substantia nigra are the earliest among the studied areas to manifest complex I loss.
(Patient AL-1B) to ~40% higher than age-matched controls in the older adults with the longest disease duration. Notably, the highest levels of excess mtDNA deletion compared to controls were found in the dopaminergic neurons of the substantia nigra. The proportion of deleted mtDNA in the patients significantly increased with age, accumulating at substantially higher levels than in controls.

FIGURE 4: mtDNA relative quantification in microdissected neurons from patients with polymerase gamma encephalopathy. Each point is a pooled sample of 5 to 15 neurons. For the purposes of comparison, a control sample has been arbitrarily set to 1. Groups are compared by Mann–Whitney U test, and p values are shown above each graph; n = number of individuals in each group. Patient neurons from all central nervous system areas examined contain significantly less (~50–60%) mtDNA than neurons of age-matched controls. The top left panel shows in addition that infants (≤1 year old) have lower neuronal mtDNA content compared to older individuals, in both the patients (infant 26% of postinfant values) and controls (infants 38% of postinfant values).
The UDS assay of mtDNA had a mean coverage of >8,500 sequence reads per amplicon in both forward and reverse directions. The overall burden of mtDNA point mutations present at a frequency >0.2% was significantly higher in patients compared to age-matched controls in both MT-HV2 (odds ratio [OR] = 3, \( p < 0.0001 \)) and MT-CO3 \( ( p = 0.025 \)). No sequence variation detectable at a frequency >0.2% was found in the highly conserved MT-CO3 region of the controls. MT-HV2 had a significantly higher mutational load than MT-CO3 in both patients and controls \( ( p > 0.0001 \) for both groups). The great majority of excess point mutations in the patients were present at low heteroplasmia levels (<2%), and their frequency decreased dramatically with increasing heteroplasmia. Overall mutational burden in MT-HV2 was significantly higher in A467T than W748S homozygous patients \( ( OR = 1.86, \ p = 0.008; \) Fig 6A), but A467T homozygous patients were also older, and this difference was no longer significant when corrected for age. In the patients, the burden of point mutations showed an apparent increase with age, although this did not reach statistical significance due to the small sample size \( ( r = 0.74, \ p = 0.09 \)). No age-dependent increase of point variants was seen in the controls \( ( r = 0.27, \ p = 0.67; \) see Fig 6B).

**Discussion**

Using a combination of clinical, neuropathological, and molecular investigations, we have defined the evolution of this devastating mitochondrial encephalopathy. The earliest feature appears to be depletion of neuronal mtDNA; this was present in the very youngest
individuals, who showed little or no morphological abnormality. Gradual loss of complex I in neurons and increasing levels of multiple mtDNA deletions and point mutations correlated with the chronic loss of neurons. Clinically, acute episodes correlated with strokelike lesions on MRI and an epileptic focus on EEG, and these in turn could be correlated with the acute focal pathological lesions that were identified throughout the cortex and in deeper structures. The correlations of clinical, pathological, and molecular features in POLG encephalopathy are summarized in Table 4.

Mutations in POLG cause a combination of mtDNA damage in the CNS including quantitative depletion, multiple deletions, and an increased burden of point mutations. Our data show that the detection and accurate quantification of depletion require studies in microdissected neurons. Quantitative studies in brain homogenate either underestimate mtDNA depletion, due to the presence of non-neuronal cells with normal mtDNA levels, or overestimate it, as in cases with severe global destruction, such as Patients AL-1B and AL-1A. Furthermore, we show that mtDNA depletion is both a universal phenomenon, that is, found in all patient neurons irrespective of type or site, and an early, but non-progressive event starting before the first year of life. That depletion remains unchanged irrespective of how long the patients lived with the disease raises the possibility that there is an absolute level below which it is not possible for the cell to survive.

Whether mtDNA depletion is present already in the ovum and/or reflects incomplete mtDNA copy number expansion during embryogenesis is unknown. Neuronal mtDNA content is known to increase from infant to adult life in normal individuals, most probably as a result of neuronal differentiation and growth. That the lowest mtDNA levels were found in the affected infant suggests that a developmental defect of mtDNA synthesis is present in patients with POLG mutations compromising this initial expansion of mtDNA. Deletions of mtDNA could not be detected in the youngest patients, but appeared in neurons of patients with longer disease duration (and who were older) at levels substantially higher than controls. Deletions affected all CNS areas studied, but showed a predilection for neurons of the substantia nigra. Nigral neurons have a known inherent predisposition for accumulating mtDNA deletions both with age and in conditions associated with neurodegeneration. One study suggested this may be related to the dopaminergic metabolism; however, irrespective of etiology, the predilection of nigral neurons for mtDNA deletions correlates with the early and severe complex I deficiency and neuronal loss in this area, suggesting that they play an important pathogenic role.

We show that UDS is an effective method and superior to cloning-based techniques for detecting and measuring somatic mtDNA mutagenesis in the brain. The finding of increased levels of mtDNA point mutations in the brain of patients with POLG mutations is novel. Although similar to previous findings in skeletal muscle, the mutational burden appeared higher in the brain, correlating with its more severe involvement in this type of POLG disease. The low individual frequency
<table>
<thead>
<tr>
<th>Feature</th>
<th>FENN</th>
<th>Diffusely Affected</th>
<th>Mildly Affected to Unaffected</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Sites</strong></td>
<td>Neocortex, hippocampus, thalamus, and cerebellar cortex</td>
<td>Spinocerebellar, dentato-olivary and dentatorubral systems, thalamus, and substantia nigra</td>
<td>All other areas studied, including non-FENN neocortex and hippocampus</td>
</tr>
<tr>
<td><strong>Clinical correlate</strong></td>
<td>Acute decompensation (strokelike episode)</td>
<td>Slow progressive decline</td>
<td>—</td>
</tr>
<tr>
<td><strong>Imaging and EEG correlate</strong></td>
<td>Acute lesions on MRI, epileptogenic foci on EEG</td>
<td>Progressive atrophy and HOD</td>
<td>—</td>
</tr>
<tr>
<td><strong>Pathological findings</strong></td>
<td>Focal, sharply demarcated lesions showing selective but incomplete neuronal loss, eosinophilic necrosis, astrocytosis, and microglial activation; vascularity may be increased</td>
<td>Diffuse progressive neuronal loss and gliosis</td>
<td>Normal morphology or mildly increased GFAP staining suggesting low-grade astrocytosis and neuronal loss</td>
</tr>
<tr>
<td>CI-deficient neurons</td>
<td>Rare</td>
<td>Significant numbers of CI-deficient neurons; proportion shows regional differences and increases significantly with age</td>
<td></td>
</tr>
<tr>
<td>CIV-deficient neurons</td>
<td>None</td>
<td>CIV-negative neurons present but significantly fewer than CI-deficient neurons; the ratio of CIV/CI-deficient neurons is significantly higher in the cerebral and cerebellar cortex than in the deep gray nuclei and brainstem</td>
<td></td>
</tr>
<tr>
<td>mtDNA depletion</td>
<td>Depletion (~40% of controls) is present from early infancy and remains stable</td>
<td></td>
<td></td>
</tr>
<tr>
<td>mtDNA deletions</td>
<td>Insufficient data</td>
<td>Not present initially; appear later and increase significantly with age mirroring the increasing number of CI- and CIV-deficient neurons</td>
<td></td>
</tr>
<tr>
<td>mtDNA point mutations</td>
<td>Not done in FENN</td>
<td>Increase with age compared to controls, reaching significantly higher levels</td>
<td></td>
</tr>
</tbody>
</table>

CI = complex I; CIV = complex IV; EEG = electroencephalogram; FENN = focal energy-dependent neuronal necrosis; GFAP = glial fibrillary acidic protein; HOD = hypertrophic olivary degeneration; MRI = magnetic resonance imaging.
of each mutation explains why they were not detected by
less sensitive techniques.12 Our data suggest a progressive
age-dependent accumulation of point mutations in the
patients, and the finding that most occur at very low
individual frequencies (<1%) suggests that these are
acquired somatic mutations caused by increased error
rate and/or impaired repair function of POLG. More-
over, the preexisting mtDNA depletion may be enhanc-
ing the process of mutagenesis by limiting the available
pool of mtDNA molecules and facilitating the clonal
expansion of those somatic mutations within cells.

Although these mutations occurred at very low fre-
quencies, a pathogenic contribution cannot be excluded.
If point mutations occur exclusively or primarily in neu-oneurons, quantifying them in brain homogenate will under-
estimate their proportion in neuronal cells. Moreover, by
occurring on a background of already substantial
mtDNA damage due to deletion and accumulating deletions, point mutations decrease further the number
of wild-type mtDNA molecules available for respiratory
complex synthesis, bringing the cells closer to the thresh-
old for respiratory deficiency.

In the face of a substantially decreased mtDNA
copy number and progressive decline in mtDNA quality,
the development of respiratory chain dysfunction is
hardly surprising. Why this selectively affects complex I
is, however, unclear. One explanation is that complex I
contains the highest number (7) of mtDNA-encoded
subunits, most of which (5) are located within the region
commonly affected by deletion. Moreover, studies suggest
that neuronal complex I is functioning close to its maxi-
mum capacity.25 It may, therefore, be the most vulnera-
table to defects of mtDNA homeostasis, including
mutations of POLG and Twinkle, the mitochondrial
helicase.26 It has been suggested that the loss of complex
I may be a protective mechanism that serves to limit the
potential damage that an inefficient respiratory chain
might cause.26 Whether complex I deficiency is a pri-
mary consequence of the disease or a secondary adaption
is a question that remains to be addressed. We found no
evidence of increased complex II activity or complex IV
loss, despite the significant degree of mtDNA damage.

Irrespective of what causes the complex I deficiency,
it appears that the ability of neurons to produce energy
is compromised. Our data suggest that this energy defi-
ciency results in 2 distinct but overlapping pathophysio-
logical processes, 1 chronic and 1 acute.

Vulnerable neuronal populations, including those in
the cerebellum, thalamus, and substantia nigra, exhibit a
substantial, time-dependent cell loss. Degeneration of the
cerebellum and its connections, including the dentato-
olivary and dentatorubral systems and dorsal column tracts,
causes a progressive spinocerebellar ataxia. Interestingly,
although cerebellar changes were significantly less pro-
nounced in the A467T homozygous patients, their ataxia
was equally severe. This may have to do with the large
proportion of complex I–negative neurons in their cerebel-
um and basal pontine nuclei, which relay the cortical
input to the cerebellum. Although these neurons have oth-
wise normal morphology, it is possible that lack of com-
plex I decreases their metabolic capacity, leading to
functional impairment. Another factor explaining the ataxia
of the A467T patients is the degeneration of the proprop-
tective dorsal spinal tracts, which is equally pronounced in
both A467T and W748S homozygous patients.

The cerebral cortex, striatum, and pons appear
more resistant to the chronic effects of complex I defi-
ciency. Neurons in these areas are able to survive, but
can still be seen as primed for injury due to their mar-
ginal energy-generating capacity. Seizures abruptly
increase neuronal energy demand, and we suggest that it
is this that precipitates the acute focal lesions in areas
such as the neocortex, hippocampus, and thalamus. Epi-
leptic activity can also explain the acute lesions in the
cerebellum. Studies have shown that epileptogenic foci
may exist within the cerebellum,27,28 but these are diffi-
cult to detect by surface recordings. Moreover, although
the role of the cerebellum in epileptogenesis remains con-
troversial, studies suggest that it may be secondarily
affected by epileptic activity elsewhere in the cortex.
Studies of perfusion using single photon emission com-
puted tomography and functional MRI show evidence of
increased cerebellar activity during clinical seizures of
cortical source and interictally.29,30 This suggests that
clinical or subclinical cortical epileptic activity may lead
to increased metabolic activity in the cerebellum, trigger-
ning acute focal lesions locally.

Neuronal energy failure would also explain why
apoptosis is not seen; apoptosis requires energy, whereas
necrosis is favored in states of severe energy deficiency.31
We suggest, therefore, that describing these lesions as
“strokelike” is inaccurate, and propose the novel descrip-
tive term focal energy-dependent neuronal necrosis.

In conclusion, this work elucidates the mechanisms
underlying neurodegeneration and clinical evolution in a
mitochondrial encephalopathy. We show how mitochon-
drial dysfunction resulting from accumulating mtDNA
damage leads to chronic neuronal loss in susceptible
regions, while at the same time it primes the brain for
acute injury triggered and propagated by epileptic seis-
ures. These findings explain the high morbidity and mor-
tality that have been associated with epilepsy in this
disorder and highlight the importance of early detection
and aggressive treatment of seizures in these patients.
Ethical Considerations
Informed consent for the autopsy was obtained from the families of patients and controls. The tissue study was approved by the Regional Ethical Committee of Western Norway (No. 2010/23).

Acknowledgment
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We thank the patients and their families for making this study possible, and Dr A. Molven and E. Fick for their valuable assistance preparing the tissue material.

Potential Conflicts of Interest
Nothing to report.

References