
Copyright:
©ERS 2015. This version is distributed under the terms of the Creative Commons Attribution Non-Commercial Licence 4.0.

DOI link to article:
http://dx.doi.org/10.1183/13993003.00891-2015

Date deposited:
27/10/2015

This work is licensed under a Creative Commons Attribution-NonCommercial 4.0 Internation License
Bile acid aspiration in people with cystic fibrosis before and after lung transplantation

To the Editor:

Cystic fibrosis (CF) is a genetic condition that is caused by abnormalities in the CF transmembrane conductance regulator (CFTR) gene. People with CF experience life-long morbidity and premature mortality, the vast majority of which is associated with lung disease.

Gastro-oesophageal reflux is known to occur frequently in children and adults with CF, and estimates of prevalence range from 55% to 90% [1]. The presence of reflux-induced cough and reported gastro-oesophageal reflux have both been associated with reduced lung function [1]. High levels of pepsin, a gastric protease, have been described in bronchoalveolar lavage (BAL) samples from children with CF [2]. People with CF are predisposed to reflux as a result of both primary and secondary mechanisms and abnormal gastrointestinal motility, with reflux of duodenal contents back into the stomach being also common [1, 3]. Consistent with these observations, high concentrations of bile acids have been described in the saliva and sputum of both adults and children with CF [1, 4].

Reflux is also a common finding in lung transplant recipients [5] and microaspiration of refluxate has been implicated in the pathogenesis of bronchiolitis obliterans syndrome (BOS) [6]. BLONDEAU et al. [1] found bile acids in 60% of BAL samples from people with CF who had undergone lung transplantation in addition to a series of other studies from the Leuven group indicating a potential role for bile acid aspiration in the pathophysiology of BOS.

We have previously shown that extraction followed by tandem mass spectrometry are necessary to document the low levels of bile acids produced by dilution effects in human BAL [7]. In pilot work, we detected bile acids in the lower airways of nine people homozygous for Phe508del with advanced CF lung disease at the time of lung transplantation [8]. We subsequently hypothesised that bile acids are present in the lower airways of people with CF before and after lung transplantation. To investigate, we used mass spectrometry to detect and identify bile acids in lower airway samples, and oesophageal pH impedance studies to provide physiological measurements of reflux.

Approval was obtained from Newcastle and North Tyneside 2 and County Durham and Tees Valley 2 Research Ethics Committees respectively. Participants underwent bilateral sequential lung transplantation for advanced CF lung disease; none had undergone antireflux surgery and all were maintained on proton pump inhibitors but none were prescribed promotility agents.

In brief, a 2×15-mL lavage was performed on explanted lungs, centrifuged and the supernatant frozen [9]. Post-transplant, a 3×60-mL BAL was performed in the same individuals at a median of 3 months following transplantation [10]. Electrospray ionisation mass spectrometry was used to detect bile acids as described in detail previously; the lower limit of detection was 0.01 μmol·L⁻¹ [7].

Combined 24-h ambulatory pH impedance was performed with the Medical Measurement System and Ohmega device (MMS Inc., Dover, NH, USA). The impedance catheter was inserted into the oesophagus using standard techniques to place the end of the catheter at the upper border of the lower oesophageal sphincter and the pH probe 5 cm above the upper border of the lower oesophageal sphincter [5].

Bile acids were detected in samples from the explanted lungs of all 19 participants with CF and in all BAL samples from the same individuals after lung transplantation (table 1). Of this group, nine out of 19 had oesophageal pH impedance studies performed post-transplant. These studies identified abnormal overall reflux in seven (78%) patients with proximal reflux in four (44%). In the seven patients with detectable reflux, it was both acid and weakly acid in six, and weakly acid alone in one patient. By comparison, bile acids, including lithocholate, were largely undetectable in BAL samples from six non-CF patients post-lung transplant (data not shown).

These results confirm that bile acids are present in the lower airways of people with advanced CF lung disease, confirming previous observations restricted to Phe508del homozygotes [8]. Longitudinal follow-up showed that bile acids continue to be detectable in the lower airways post-lung transplantation.

Copyright 2015 by the European Respiratory Society.
Furthermore, we found abnormal levels of reflux in most people with CF tested post-transplant. This rare longitudinal information indicates that reflux and aspiration are not simply a function of factors associated with chronic lung disease; we demonstrate persistence post-transplantation, where cough is attenuated while thoracic mechanical changes caused by advanced lung disease are corrected.

We used mass spectrometric bile assays, suitable for the low levels found in BAL samples, providing specific information about individual bile acids [7]. There are no studies characterising individual aspirated bile acids in the CF lung that we are aware of. We measured median lithocholate levels of 0.2 and 0.4 µmol·L\(^{-1}\) pre- and post-lung transplantation, respectively. Lithocholic acid was recently detected following direct tracheal suction at a mean±SEM concentration of 2.8±0.6 µmol·L\(^{-1}\) in critically unwell patients with ventilator-associated pneumonia [11]. Elevated levels of bile acids were associated with lung injury and were raised compared to ventilated patients with no pneumonia. Furthermore, Wu et al. [11] showed that chenodeoxycholic acid challenge of alveolar epithelial cells led to interleukin-8 generation.

Lithocholate is normally a minor component of circulating bile acids, produced by specific gut bacteria that deconjugate and dehydroxylate the primary bile acid chenodeoxycholate. Bile acid homeostasis and metabolism may be disordered in CF [12]. CF is a multisystem disorder that involves both the respiratory and gastrointestinal tracts. Duodeno-gastro-oesophageal reflux and subsequent microaspiration of bile acids may be an under-recognised contributor to airway injury in CF lung disease [13]. Treatments for reflux exist and a very limited literature has advocated fundoplication in CF. In a recent small open study of adult CF patients there was a fall in cough, and exacerbation events reduced by 50% post-fundoplication [13]. Fundoplication in children with CF has also been undertaken but results have been varied [14].

It is important that we acknowledge the limitations of our study. For technical reasons, a small-volume lavage was performed in the noninflated resected lungs compared to the 180-mL BAL used to sample allografts. There is still no entirely satisfactory method of determining the dilution factor during BAL.

Post-transplantation, people with CF have comparable levels of BOS to patients transplanted for other indications. However, the post-transplantation CF population is heterogeneous and some individuals develop BOS with an aggressive onset; this is now recognised as a complex, alloimmune and non-alloimmune disorder.

---

**TABLE 1** Brief clinical details and bile acid concentrations pre- and post-lung transplantation measured by electrospray ionisation mass spectrometry

<table>
<thead>
<tr>
<th>Patient</th>
<th>Age(^{a}) years</th>
<th>Sex</th>
<th>pH im.</th>
<th>Explanted lung samples(^{b}) µmol·L(^{-1})</th>
<th>Post-transplant bronchoalveolar \lavage samples(^{b}) µmol·L(^{-1})</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>32</td>
<td>Female</td>
<td>+</td>
<td>0.01</td>
<td>ND</td>
</tr>
<tr>
<td>2</td>
<td>25</td>
<td>Female</td>
<td>+</td>
<td>0.03</td>
<td>0.01</td>
</tr>
<tr>
<td>3</td>
<td>23</td>
<td>Male</td>
<td>–</td>
<td>0.03</td>
<td>0.03</td>
</tr>
<tr>
<td>4</td>
<td>39</td>
<td>Male</td>
<td>–</td>
<td>0.23</td>
<td>0.05</td>
</tr>
<tr>
<td>5</td>
<td>27</td>
<td>Male</td>
<td>–</td>
<td>0.09</td>
<td>0.06</td>
</tr>
<tr>
<td>6</td>
<td>29</td>
<td>Male</td>
<td>+</td>
<td>0.25</td>
<td>0.04</td>
</tr>
<tr>
<td>7</td>
<td>30</td>
<td>Female</td>
<td>+</td>
<td>0.10</td>
<td>0.01</td>
</tr>
<tr>
<td>8</td>
<td>23</td>
<td>Female</td>
<td>+</td>
<td>0.06</td>
<td>0.07</td>
</tr>
<tr>
<td>9</td>
<td>46</td>
<td>Male</td>
<td>+</td>
<td>0.17</td>
<td>0.18</td>
</tr>
<tr>
<td>10</td>
<td>29</td>
<td>Female</td>
<td>+</td>
<td>0.08</td>
<td>0.04</td>
</tr>
<tr>
<td>11</td>
<td>22</td>
<td>Female</td>
<td>+</td>
<td>0.04</td>
<td>0.03</td>
</tr>
<tr>
<td>12</td>
<td>20</td>
<td>Female</td>
<td>–</td>
<td>0.16</td>
<td>0.11</td>
</tr>
<tr>
<td>13</td>
<td>29</td>
<td>Female</td>
<td>+</td>
<td>0.06</td>
<td>0.05</td>
</tr>
<tr>
<td>14</td>
<td>48</td>
<td>Female</td>
<td>–</td>
<td>0.03</td>
<td>0.04</td>
</tr>
<tr>
<td>15</td>
<td>22</td>
<td>Male</td>
<td>–</td>
<td>0.10</td>
<td>0.07</td>
</tr>
<tr>
<td>16</td>
<td>24</td>
<td>Male</td>
<td>–</td>
<td>0.03</td>
<td>0.03</td>
</tr>
<tr>
<td>17</td>
<td>21</td>
<td>Male</td>
<td>–</td>
<td>0.64</td>
<td>0.64</td>
</tr>
<tr>
<td>18</td>
<td>33</td>
<td>Female</td>
<td>–</td>
<td>0.08</td>
<td>0.02</td>
</tr>
<tr>
<td>19</td>
<td>29</td>
<td>Male</td>
<td>–</td>
<td>0.72</td>
<td>0.19</td>
</tr>
<tr>
<td>Median</td>
<td></td>
<td></td>
<td>0.08</td>
<td>0.04</td>
<td>0.02</td>
</tr>
</tbody>
</table>

\(^{a}\): pH impedance; G-DHC: glycodehydroxycholanoate (deoxycholate and isomers, glycine conjugate); G-THC: glycotrihydroxycholanoate (cholate and isomers, glycine conjugate); T-DHC: taurodihydroxycholanoate (deoxycholate and isomers, taurine conjugate); T-THC: taurotrihydroxycholanoate (cholate and isomers, taurine conjugate); Litho.: unconjugated lithocholate; +: performed; –: not performed; ND: not detected [less than lower limit of detection 0.01 µmol·L\(^{-1}\)]. \(^{b}\): at time of transplant [complete years only]; \(^{c}\): raw data of bile acids detected in 180-mL bronchoalveolar lavages. \(^{d}\): for bile acids where the level was below the limit of detection, the median was calculated after nondetectable samples were assigned an arbitrary low value of 0.01 µmol·L\(^{-1}\).
response to injury. Colonisation of CF allografts with Pseudomonas aeruginosa has been associated with bile aspiration and neutrophilic airway inflammation by the Leuven group [15]. In turn, infection and inflammation, associated with bile aspiration [11, 15], have been linked to BOS and death in 260 lung transplant recipients in a recent series [16]. Further studies are therefore required in greater numbers of patients. There is potential that outcomes of lung transplantation in CF, already comparable with other transplant recipients in a recent series [16], could be further improved if aspiration injury were reliably detected and treated.

In conclusion, we have shown that bile acids measured by tandem mass spectrometry are detectable in the lower airway in advanced CF lung disease and that this potential source of injury persists after lung transplantation. Interactions between gastrointestinal and lung pathophysiologies may contribute to overall lung damage and this "aerodigestive" concept may have therapeutic implications in a disease where available treatments are otherwise limited.

Bile acids are detectable in the lower airway in advanced CF lung disease and persist after lung transplantation

Malcolm Brodlie1,2, Ali Aseeri1,3, Jim L. Lordan1,4, Andrew G.N. Robertson5, Michael C. McKean2, Paul A. Corris1,4, S. Michael Griffin1, Nigel J. Manning6, Jeffrey P. Pearson1 and Christopher Ward1

1Institute of Cellular Medicine, Newcastle University, Newcastle upon Tyne, UK. 2Department of Paediatric Respiratory Medicine, Great North Children’s Hospital, Newcastle upon Tyne Hospitals NHS Foundation Trust, Newcastle upon Tyne, UK. 3Institute for Cell and Molecular Bioscience, Newcastle University, Newcastle upon Tyne, UK. 4Institute of Transplantation, Freeman Hospital, Newcastle upon Tyne Hospitals NHS Foundation Trust, Newcastle upon Tyne, UK. 5Northern Oesophago-Gastric Unit, Royal Victoria Infirmary, Newcastle upon Tyne Hospitals NHS Foundation Trust, Newcastle upon Tyne, UK. 6Department of Clinical Chemistry, Sheffield Children’s Hospital, The Sheffield Children’s NHS Trust, Sheffield, UK.

Correspondence: Chris Ward, Room 1.072, Catherine Cookson Building, Medical School, Newcastle University, Framlington Place, Newcastle upon Tyne, NE2 4HH, UK. E-mail: chris.ward@ncl.ac.uk

Received: March 31 2015 | Accepted after revision: Aug 12 2015

Support statement: Funding for this work was gratefully received from a Saudi Arabian government studentship (A. Aseeri, J.P. Pearson and C. Ward), Medical Research Council (MRC)/CF Trust Clinical Research Training and Clinician Scientist Fellowships (M. Brodlie), an MRC Project Grant (C. Ward and P.A. Corris), the Newcastle upon Tyne Hospitals NHS Foundation Trust Special Trustees (M.C. McKean, M. Brodlie, C. Ward and J.L. Lordan) and the British Lung Foundation Trevor Clay Memorial Award (C. Ward, J.P. Pearson and A.G.N. Robinson). Funding information for this article has been deposited with FundRef.

Conflict of interest: None declared.

Acknowledgements: We gratefully acknowledge the help of the patients involved in this study along with all members of the wider cardiothoracic transplantation team at the Freeman Hospital.

References

