

Jose S, Quinn K, Dunn D, Cox A, Sabin C, Fidler S, Fisher M, Gazzard B, Gilson R, Gompels M, Hay P, Johnson M, Kegg S, Leen C, Martin F, Nelson M, Palfreeman A, Post F, Pritchard J, Sachikonye M, Schwenk A, Tariq A, Walsh J, Hill T, Jose S, Phillips A, Sabin C, Thornton A, Dunn D, Glabay A, Fisher M, Perry N, Tilbury S, Youssef E, Churchill D, Gazzard B, Nelson M, Everett R, Asboe D, Mandalia S, Post F, Korat H, Taylor C, Gleisner Z, Ibrahim F, Campbell L, Gilson R, Brima N, Williams I, Johnson M, Youle M, Lampe F, Smith C, Tsintas R, Chaloner C, Hutchinson S, Phillips A, Hill T, Thornton A, Huntington S, Walsh J, Mackie N, Winston A, Weber J, Ramzan F, Carder M, Orkin C, Lynch J, Hand J, de Souza C, Anderson J, Munshi S, Ainsworth J, Schwenk A, Miller S, Wood C, Leen C, Wilson A, Morris S, Gompels M, Allan S, Palfreeman A, Memon K, Lewszuk A, Chadwick D, Cope E, Gibson J, Kegg S, Main P, Mitchell, Hunter, Hay P, Dhillon M, Martin F, Russell-Sharp S, Allan S, Harte A, Clay S, Tariq A, Spencer H, Jones R, Pritchard J, Cumming S, Atkinson C, Delpech V, Sachikony M, Aitken C, Asboe D, Pozniak A, Cane P, Chadwick D, Churchill D, Clark D, Collins S, Delpech V, Douthwaite S, Fearnhill E, Porter K, Tostevin A, White E, Fraser C, Geretti AM, Hale A, Hue S, Kaye S, Kellam P, Lazarus L, Leigh-Brown A, Mbisa T, Mackie N, Moses S, Orkin C, Nastouli E, Pillay D, Smit E, Templeton K, Tilston P, Webster D, Williams I, Zhang H, Greatorex J, O'Shea S, Mullen J, Cox A, Tandy R, Fawcett T, Hopkins M, Ashton L, Booth C, Garcia-Diaz A, Shepherd J, Schmid ML, Payne B, Pereira S, Hubb J, Kirk S, Gunson R, Bradley-Stewart A. [Virological failure and development of new resistance mutations according to CD4 count at combination antiretroviral therapy initiation](#). *HIV Medicine* 2016, 17(5), 368-372.



This work is licensed under a [Creative Commons Attribution 4.0 International License](#)

**Copyright:**

© 2015 The Authors. *HIV Medicine* published by John Wiley & Sons Ltd on behalf of British HIV Association.

This is an open access article under the terms of the [Creative Commons Attribution](#) License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

**DOI link to article:**

<http://dx.doi.org/10.1111/hiv.12302>

**Date deposited:**

03/04/2017



This work is licensed under a [Creative Commons Attribution 4.0 International License](#)

# Virological failure and development of new resistance mutations according to CD4 count at combination antiretroviral therapy initiation

S Jose,<sup>1</sup> K Quinn,<sup>2</sup> D Dunn,<sup>3</sup> A Cox,<sup>4</sup> C Sabin,<sup>1</sup> S Fidler,<sup>2</sup> on behalf of the UK CHIC and UK HDRD Steering Committees<sup>†</sup>  
<sup>1</sup>Research Department of Infection and Population Health, UCL, London, UK, <sup>2</sup>Department of Medicine, Imperial College London, London, UK, <sup>3</sup>Medical Research Council Clinical Trials Unit at UCL, London, UK and <sup>4</sup>Infection and Immunity, Imperial College Healthcare NHS Trust, London, UK

## Objectives

No randomized controlled trials have yet reported an individual patient benefit of initiating combination antiretroviral therapy (cART) at CD4 counts > 350 cells/ $\mu$ L. It is hypothesized that earlier initiation of cART in asymptomatic and otherwise healthy individuals may lead to poorer adherence and subsequently higher rates of resistance development.

## Methods

In a large cohort of HIV-positive individuals, we investigated the emergence of new resistance mutations upon virological treatment failure according to the CD4 count at the initiation of cART.

## Results

Of 7918 included individuals, 6514 (82.3%), 996 (12.6%) and 408 (5.2%) started cART with a CD4 count  $\leq$  350, 351–499 and  $\geq$  500 cells/ $\mu$ L, respectively. Virological rebound occurred while on cART in 488 (7.5%), 46 (4.6%) and 30 (7.4%) with a baseline CD4 count  $\leq$  350, 351–499 and  $\geq$  500 cells/ $\mu$ L, respectively. Only four (13.0%) individuals with a baseline CD4 count > 350 cells/ $\mu$ L in receipt of a resistance test at viral load rebound were found to have developed new resistance mutations. This compared to 107 (41.2%) of those with virological failure who had initiated cART with a CD4 count < 350 cells/ $\mu$ L.

## Conclusions

We found no evidence of increased rates of resistance development when cART was initiated at CD4 counts above 350 cells/ $\mu$ L.

**Keywords:** antiretroviral therapy, CD4 count, HIV resistance, virological failure

Accepted 1 July 2015

## Introduction

Although morbidity and mortality benefits of starting combination antiretroviral therapy (cART) at CD4 counts > 350 cells/ $\mu$ L have been reported in cohort studies [1,2], there is little randomized evidence on the individual risk–benefit ratio of initiating combination antiretroviral therapy (cART) at higher CD4 counts [3]. The

randomized controlled Strategic Timing of AntiRetroviral Treatment (START) trial has recently investigated the optimal timing of cART initiation in order to improve morbidity and mortality outcomes in HIV-positive individuals [4]. Nevertheless, there have already been changes to national and international HIV treatment guidelines [5,6], largely driven by the impact of cART on viral transmission [7] and a pragmatic approach to cART roll out programmes.

As adherence to cART has been associated with perceived “need” for treatment [8], there is concern that a recommendation to start cART at higher CD4 counts may be met with patients’ ambivalence to cART, leading to suboptimal adherence and antiretroviral resistance. However, there are no data reported in support of this hypothesis to date [9,10]. The START trial will investigate

Correspondence: Sophie Jose, Research Department of Infection and Population Health UCL, Royal Free Campus, London, NW3 2PF, UK. Tel: 02077940500, ext. 36763; e-mail: sophie.jose@ucl.ac.uk

<sup>†</sup>The UK CHIC and UK HDRD Steering Committees members are listed in Appendix 1.

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

antiretroviral resistance development as a secondary endpoint, and will report these findings after 2016.

Previously, we reported on laboratory-defined adverse events (LDAEs) according to the CD4 count at initiation of cART [11]. We now describe rates of new antiretroviral resistance mutations in those experiencing virological failure, according to the CD4 count at treatment initiation.

## Methods

The UK Collaborative HIV Cohort (CHIC) Study collates routinely collected clinical data on HIV-positive individuals accessing care across several centres in the UK. The study was approved by a multicentre research ethics committee and by local ethics committees and does not require informed consent. Similarly, the UK HIV Drug Resistance Database (HDRD) collates results of routine resistance testing of HIV-positive individuals accessing care. A combined UK CHIC – UK HDRD dataset was used for analysis. Individuals were included who initiated cART ( $\geq 3$  antiretroviral drugs) between 2000 and 2011 with a baseline CD4 count and resistance test result available, and who achieved an undetectable viral load on cART. Pregnant women were excluded.

Virological rebound was defined as the first occurrence of two consecutive viral loads  $> 400$  HIV-1 RNA copies/mL following an undetectable viral load. Virological rebound occurring while on cART was classed as treatment failure and evaluated for resistance development. Virological rebound that occurred immediately following a treatment discontinuation was not evaluated for resistance development. The earliest resistance test result available up to 1 month before or 6 months after the date of virological failure was used. We defined new resistance as the presence of any new major resistance mutation [12] not present in the baseline genotype, stratifying results according to the CD4 count at cART initiation ( $\leq 350$ , 351–499 and  $\geq 500$  cells/ $\mu$ L).

## Results

In total, 9288 people had initiated cART since 2000 and had a baseline CD4 count and resistance test result available. Of these, 8445 (91%) achieved an undetectable viral load. A further 527 were excluded due to pregnancy. Of 7918 included, 6514 (82.3%), 996 (12.6%) and 408 (5.2%) started cART with a CD4 count  $\leq 350$ , 351–99 and  $\geq 500$  cells/ $\mu$ L, respectively.

Those with a baseline CD4 count 351–499 or  $\geq 500$  cells/ $\mu$ L were more likely to be men who have sex with men (MSM), of white ethnicity, co-infected with HCV and

having started cART in a later calendar year than those with a baseline CD4 count  $\leq 350$  cells/ $\mu$ L ( $P < 0.0001$ ). Those with a CD4 count  $\geq 500$  cells/ $\mu$ L were more likely to start a ritonavir-boosted protease inhibitor (PI/r)-based regimen (43.9 *vs.* 29.3% 351–499 cells/ $\mu$ L and 25.8%  $< 350$  cells/ $\mu$ L) and less likely to start a nonnucleoside reverse transcriptase inhibitor (NNRTI)-based regimen (52.0 *vs.* 64.7% and 69.5%, respectively). At baseline, 255 (4.0%), 26 (2.7%) and 23 (5.9%) of those with CD4 count  $\leq 350$ , 351–499 and  $\geq 500$  cells/ $\mu$ L were not fully susceptible to  $\geq 1$  drug in their cART regimen ( $P = 0.02$ ).

Virological rebound occurred in 806 (10.2%) individuals, of whom 564 were receiving cART at the time of rebound: 488 (7.5%), 46 (4.6%) and 30 (7.4%) with a baseline CD4 count  $\leq 350$ , 351–499 and  $\geq 500$  cells/ $\mu$ L, respectively. A higher proportion of individuals with a baseline CD4 count  $\leq 350$  cells/ $\mu$ L received a resistance test at virological failure (Table 1). By the time of virological failure, 30.0% had changed their cART regimen class, and 38.5, 54.4 and 53.3% were on PI/r-based, and 45.3, 34.8 and 40.0% were on a NNRTI-based regimens ( $\leq 350$ , 351–499 and  $\geq 500$  cells/ $\mu$ L, respectively). The median (interquartile range) viral load was 3.9 (3.1, 4.8), 4.3 (3.4, 5.0) and 3.6 (3.4, 4.5)  $\log_{10}$ copies/mL and 18.9, 15.2 and 13.3% had a viral load  $< 1000$  copies/mL ( $\leq 350$ , 351–499 and  $\geq 500$  cells/ $\mu$ L, respectively). Only four of 30 with a baseline CD4 count  $> 350$  cells/ $\mu$ L had evidence of new resistance mutations at viral load rebound. All mutations conferred resistance to either the nucleoside reverse transcriptase inhibitor (NRTI) or NNRTI class of antiretroviral drugs. New PI, NNRTI and NRTI resistance mutations were found in 3.3, 26.5 and 27.3% of those with a baseline CD4 count  $\leq 350$  cells/ $\mu$ L and 18.1% had resistance to more than one class of antiretroviral drug.

## Discussion

Our results do not demonstrate an increased risk of virological failure on cART when therapy is initiated at CD4 counts  $> 350$  cells/ $\mu$ L. In fact, there appeared to be a reduced risk of developing a major resistance mutation when cART was initiated at CD4 counts  $> 350$  cells/ $\mu$ L. Greater use of PI/r-based regimens with a higher genetic barrier to resistance in this group may explain the effect of seeing fewer resistance mutations emerge when virological failure occurred [13]. However, our findings are consistent with other studies [9,10].

Higher rates of virological rebound were observed in the group that started cART at CD4 counts above 500 cells/ $\mu$ L. The majority of rebounds in this group occurred following treatment discontinuation, with the proportion

of patients experiencing rebound while reportedly still receiving cART being similar across groups. Reasons for treatment discontinuation in this group are unknown; however, it is possible that some of these individuals were enrolled in trials of treatment interruption strategies that were undertaken during this time period [14]. However, this trend towards more treatment interruption in those starting cART with a high CD4 count raises some concerns. Depending on the specific drugs included in the regimen as well as the timing of stopping, there is the potential for viral replication to occur in the presence of sub-optimal levels of cART following treatment discontinuation, thus leading to selection of drug-resistant virus, particularly for regimens containing NNRTIs [15]. Of those who experienced virological rebound following treatment interruption [45 (11%)  $\geq$  500, 42 (4.2%) 351–499, 155 (2.4%)  $\leq$  350 cells/ $\mu$ L], nine (20%), nine (21%) and 62 (40%) had a resistance test ( $>$  500, 351–499 and  $<$  350 cells/ $\mu$ L, respectively) with only one (11.1%) person having new resistance mutations detected for CD4 counts 351–499 cells/ $\mu$ L and 15 (24.2%) for CD4 counts  $\leq$  350 cells/ $\mu$ L.

Fewer resistance tests appeared to be undertaken in individuals experiencing virological failure who had initiated treatment with CD4 counts  $>$  350 cells/ $\mu$ L, raising concerns that resistance mutations may be missed in this group. However, this may be a chance finding and reasons for a lower rate of testing in this group are unclear; lower viral load at virological failure did not appear to explain this and no significant predictors of resistance testing at virological failure were found in any CD4 strata.

Our analyses are limited because reasons for starting cART at high CD4 counts outside current national guidelines are not known. Caucasians and MSM were over-represented amongst those starting therapy with CD4 counts  $>$  350 cells/ $\mu$ L, indicating that subgroups traditionally presenting to HIV services earlier in the United

Kingdom tend to initiate treatment earlier [16]. This may indicate an underlying selection bias amongst those starting therapy early, as native English speakers and UK nationals with greater access to healthcare and of potentially higher educational status may opt to start therapy earlier. However, repeating our analysis including only MSM, we saw similar patterns by CD4 count strata, with 35.1, 12.5 and 11.1% of those with CD4 counts  $<$  350, 351–499 and  $>$  500 cells/ $\mu$ L (respectively) having new resistance mutations when tested at virological rebound. Furthermore, in this observational setting, those who have been motivated to start cART at higher CD4 counts may be more likely to have better adherence to treatment and may therefore be less likely to either experience virological rebound or develop resistance to antiretrovirals.

Despite certain limitations, we have not found evidence of an increased risk of resistance development at virological failure amongst people initiating cART at CD4 counts  $>$  350 cells/ $\mu$ L.

## Acknowledgements

**Funding:** This work was supported by the Medical Research Council, UK [grant numbers G0000199, G0600337, G0900274 and M004236]. The views expressed in this manuscript are those of the researchers and not necessarily those of the MRC.

**Conflicts of interest:** Sophie Jose, Killian Quinn, Alison Cox and David Dunn have no conflicts of interest to report.

Caroline Sabin has received honoraria participation in Data Safety and Monitoring Boards and Advisory Boards, for preparation of educational materials and for membership of speaker panels for Gilead Sciences, Bristol-Myers Squibb, ViiV Healthcare/GlaxoSmithKline, Janssen-Cilag and Abbvie. Sarah Fidler has worked in research collaborations with MSD and GlaxoSmithKline and as a speaker for ViiV Healthcare, Janssen-Cilag and MSD.

**Table 1** Virological failure and development of resistance according to CD4 count at combination antiretroviral therapy (cART) initiation

Baseline CD4 count, cells/ $\mu$ L	N	Virological rebound defined, n (%)	Virological rebound occurring on treatment, n (%)	Resistance test, n (%)	$\geq$ 1 new resistance mutation, n (%)	PI resistance, n (%)	NNRTI resistance, n (%)	NRTI resistance, n (%)
$\leq$ 350	6514	643 (9.9)	488 (7.5)	260 (53.3)	107 (41.2)	8 (3.1)	69 (26.5)	71 (27.3)
351–499	996	88 (8.8)	46 (4.6)	20 (43.5)	3 (15.0)	0 (0.0)	3 (15.0)	2 (10.0)
$\geq$ 500	408	75 (18.4)	30 (7.4)	10 (33.3)	1 (10.0)	0 (0.0)	1 (10.0)	1 (10.0)
Total	7918	806 (10.2)	564 (7.1)	290 (51.4)	111 (38.3)	8 (1.4)	73 (25.2)	74 (25.5)
P-value*	$<$ 0.0001		0.005	0.056	0.012			

PI, protease inhibitor; NNRTI, non-nucleoside reverse transcriptase inhibitor, NRTI, nucleoside reverse transcriptase inhibitor.

940 (11.9%) people had at least one major resistance mutation at baseline; of these, 778 (11.9%), 113 (11.4%) and 49 (12.0%) had baseline CD4 count  $\leq$  350, 351–499 and  $\geq$  500 cells/ $\mu$ L ( $P = 0.86$ ).

\*All  $P$ -values based on a chi squared test.

**Note**

\* Coordinating Centre.

**Appendix (1)****UK CHIC Study**

Steering Committee: Jonathan Ainsworth, Sris Allan, Jane Anderson, Abdel Babiker, David Chadwick, Valerie Delpech, David Dunn, Martin Fisher, Brian Gazzard, Richard Gilson, Mark Gompels, Phillip Hay, Teresa Hill, Margaret Johnson, Sophie Jose, Stephen Kegg, Clifford Leen, Fabiola Martin, Mark Nelson, Chloe Orkin, Adrian Palfreeman, Andrew Phillips, Deenan Pillay, Frank Post, Jillian Pritchard, Caroline Sabin, Memory Sachikonye, Achim Schwenk, Anjum Tariq, John Walsh.

Central Co-ordination: University College London (Teresa Hill, Sophie Jose, Andrew Phillips, Caroline Sabin, Alicia Thornton); Medical Research Council Clinical Trials Unit at UCL (MRC CTU at UCL), London (David Dunn, Adam Glabay).

Participating Centres: Brighton and Sussex University Hospitals NHS Trust (M Fisher, N Perry, S Tilbury, E Youssef, D Churchill); Chelsea and Westminster Hospital NHS Foundation Trust, London (B Gazzard, M Nelson, R Everett, D Asboe, S Mandalia); King's College Hospital NHS Foundation Trust, London (F Post, H Korat, C Taylor, Z Gleisner, F Ibrahim, L Campbell); Mortimer Market Centre, University College London (R Gilson, N Brima, I Williams); Royal Free NHS Foundation Trust/University College London (M Johnson, M Youle, F Lampe, C Smith, R Tsintas, C Chaloner, S Hutchinson, C Sabin, A Phillips, T Hill, S Jose, A Thornton, S Huntington); Imperial College Healthcare NHS Trust, London (J Walsh, N Mackie, A Winston, J Weber, F Ramzan, M Carder); Barts and The London NHS Trust, London (C Orkin, J Lynch, J Hand, C de Souza); Homerton University Hospital NHS Trust, London (J Anderson, S Munshi); North Middlesex University Hospital NHS Trust, London (J Ainsworth, A Schwenk, S Miller, C Wood); The Lothian University Hospitals NHS Trust, Edinburgh (C Leen, A Wilson, S Morris); North Bristol NHS Trust (M Gompels, S Allan); Leicester, University Hospitals of Leicester NHS Trust (A Palfreeman, K Memon, A Lewszuk); Middlesbrough, South Tees Hospitals NHS Foundation Trust (D Chadwick, E Cope, J Gibson); Woolwich, Lewisham and Greenwich NHS Trust (S Kegg, P Main, Dr Mitchell, Dr Hunter), St George's Healthcare NHS Trust (P Hay, M Dhillon); York Teaching Hospital NHS Foundation Trust (F Martin, S Russell-Sharpe); Coventry, University Hospitals Coventry and

Warwickshire NHS Trust (S Allan, A Harte, S Clay); Wolverhampton, The Royal Wolverhampton Hospitals NHS Trust (A Tariq, H Spencer, R Jones); Chertsey, Ashford and St Peter's Hospitals NHS Foundation Trust (J Pritchard, S Cumming, C Atkinson); Public Health England, London (V Delpech); UK Community Advisory Board (M Sachikonye).

**UK HDRD**

Steering Committee: Celia Aitken (Gartnavel General Hospital, Glasgow); David Asboe, Anton Pozniak (Chelsea & Westminster Hospital, London); Patricia Cane (Public Health England, Porton Down); David Chadwick (South Tees Hospitals NHS Trust, Middlesbrough); Duncan Churchill (Brighton and Sussex University Hospitals NHS Trust); Duncan Clark (St Bartholomew's and The London NHS Trust); Simon Collins (HIV i-Base, London); Valerie Delpech (Centre for Infections, Public Health England); Samuel Douthwaite (Guy's and St. Thomas' NHS Foundation Trust, London); David Dunn, Esther Fearnhill, Kholoud Porter, Anna Tostevin, Ellen White (MRC Clinical Trials Unit at UCL, London)\*; Christophe Fraser (Imperial College London); Anna Maria Geretti (Institute of Infection and Global Health, University of Liverpool); Antony Hale (Leeds Teaching Hospitals NHS Trust); Stéphane Hué (University College London); Steve Kaye (Imperial College, London); Paul Kellam (Wellcome Trust Sanger Institute & University College London Medical School); Linda Lazarus (Expert Advisory Group on AIDS Secretariat, Public Health England); Andrew Leigh-Brown (University of Edinburgh); Tamyo Mbisa (Virus Reference Department, Public Health England); Nicola Mackie (Imperial NHS Trust, London); Samuel Moses (King's College Hospital, London); Chloe Orkin (St. Bartholomew's Hospital, London); Eleni Nastouli, Deenan Pillay, Andrew Phillips, Caroline Sabin (University College London Medical School, London); Erasmus Smit (Public Health England, Birmingham Heartlands Hospital); Kate Templeton (Royal Infirmary of Edinburgh); Peter Tilston (Manchester Royal Infirmary); Daniel Webster (Royal Free NHS Trust, London); Ian Williams (Mortimer Market Centre, London); Hongyi Zhang (Addenbrooke's Hospital, Cambridge).

Centres contributing data: Clinical Microbiology and Public Health Laboratory, Addenbrooke's Hospital, Cambridge (Jane Greatorex); Guy's and St. Thomas' NHS Foundation Trust, London (Siobhan O'Shea, Jane Mullen); PHE – Public Health Laboratory, Birmingham Heartlands Hospital, Birmingham (Erasmus Smit); PHE – Virus Reference Department, London (Tamyo Mbisa); Imperial Col-

lege Health NHS Trust, London (Alison Cox); King's College Hospital, London (Richard Tandy); Medical Microbiology Laboratory, Leeds Teaching Hospitals NHS Trust (Tracy Fawcett); Specialist Virology Centre, Liverpool (Mark Hopkins, Lynn Ashton); Department of Clinical Virology, Manchester Royal Infirmary, Manchester (Peter Tilston); Department of Virology, Royal Free Hospital, London (Claire Booth, Ana Garcia-Diaz); Edinburgh Specialist Virology Centre, Royal Infirmary of Edinburgh (Jill Shepherd); Department of Infection & Tropical Medicine, Royal Victoria Infirmary, Newcastle (Matthias L Schmid, Brendan Payne); South Tees Hospitals NHS Trust, Middlesbrough (David Chadwick); Department of Virology, St Bartholomew's and The London NHS Trust (Spiro Pereira, Jonathan Hubb); Molecular Diagnostic Unit, Imperial College, London (Steve Kaye); University College London Hospitals (Stuart Kirk); West of Scotland Specialist Virology Laboratory, Gartnavel, Glasgow (Rory Gunson, Amanda Bradley-Stewart, Celia Aitken).

## References

- 1 Kitahata MM, Gange SJ, Abraham AG *et al.* Effect of early versus deferred antiretroviral therapy for HIV on survival. *N Engl J Med* 2009; **360**: 1815–1826.
- 2 Sterne JA, May M, Costagliola D *et al.* Timing of initiation of antiretroviral therapy in AIDS-free HIV-1-infected patients: a collaborative analysis of 18 HIV cohort studies. *Lancet* 2009; **373**: 1352–1363.
- 3 Sabin CAC, David A, Collins S, Schechter M. Rating evidence in treatment guidelines: a case example of when to initiate combination antiretroviral therapy (cART) in HIV-positive asymptomatic persons. *AIDS* 2013; **27**: 1839–1846.
- 4 Babiker AG, Emery S, Fatkenheuer G *et al.* Considerations in the rationale, design and methods of the Strategic Timing of AntiRetroviral Treatment (START) study. *Clin Trials* 2013; **10** (1 Suppl): S5–S36.
- 5 Thompson MA, Aberg JA, Hoy JF *et al.* Antiretroviral treatment of adult HIV infection: 2012 recommendations of the International Antiviral Society-USA panel. *JAMA* 2012; **308**: 387–402.
- 6 World Health Organization. Consolidated Guidelines on the Use of Antiretroviral Drugs for Treating and Preventing HIV Infection. Recommendations for a Public Health Approach. <http://www.who.int/hiv/pub/guidelines/arv2013/en/index.html> (accessed August 07, 2013). 2013.
- 7 Cohen MS, Chen YQ, McCauley M *et al.* Prevention of HIV-1 infection with early antiretroviral therapy. *N Engl J Med* 2011; **365**: 493–505.
- 8 Horne R, Buick D, Fisher M, Leake H, Cooper V, Weinman J. Doubts about necessity and concerns about adverse effects: identifying the types of beliefs that are associated with non-adherence to HAART. *Int J STD AIDS* 2004; **15**: 38–44.
- 9 Uy J, Armon C, Buchacz K, Wood K, Brooks JT. Initiation of HAART at higher CD4 cell counts is associated with a lower frequency of antiretroviral drug resistance mutations at virologic failure. *J Acquir Immune Defic Syndr* 2009; **51**: 450–453.
- 10 Lodi S, Phillips A, Fidler S *et al.* Role of HIV infection duration and CD4 cell level at initiation of combination antiretroviral therapy on risk of failure. *PLoS ONE* 2013; **8**: e75608.
- 11 Jose S, Quinn K, Hill T *et al.* Laboratory adverse events and discontinuation of therapy according to CD4+ cell count at the start of antiretroviral therapy. *AIDS* 2014; **28**: 1333–1339.
- 12 Wensing AM, Calvez V, Gunthard HF *et al.* 2014 Update of the drug resistance mutations in HIV-1. *Top Antivir Med* 2014; **22**: 642–650.
- 13 Gardner EM, Burman WJ, Steiner JF, Anderson PL, Bangsberg DR. Antiretroviral medication adherence and the development of class-specific antiretroviral resistance. *AIDS* 2009; **23**: 1035–1046.
- 14 Fidler S, Porter K, Ewings F *et al.* Short-course antiretroviral therapy in primary HIV infection. *N Engl J Med* 2013; **368**: 207–217.
- 15 Arnedo-Valero M, Garcia F, Gil C *et al.* Risk of selecting de novo drug-resistance mutations during structured treatment interruptions in patients with chronic HIV infection. *Clin Infect Dis* 2005; **41**: 883–890.
- 16 Aghaizu A, Brown A, Nardone A, Gill O, Delpech V. HIV in the United Kingdom 2013 Report: data to end 2012. Public Health England. [http://www.hpa.org.uk/webc/HPAwebFile/HPAweb\\_C/1317140300680](http://www.hpa.org.uk/webc/HPAwebFile/HPAweb_C/1317140300680) (accessed 04 April 2014).